Post-Graduate Course: Lung innate immunity

Emerging roles of ILCs in mucosal barrier

Seoul National University College of Medicine

Hye Young Kim

1. Introduction

Innate lymphoid cells (ILCs) are widely distributed in the body from hematopoietic organs such as the BM to secondary lymphoid tissues and non-lymphoid tissues. Among them, intestine, skin, lungs, and adipose tissues are relatively more enriched with ILCs, which account for $\sim 2.5\%$ of hematopoietic (i.e. CD45⁺) cells in the intestinal lamina propria, $\sim 3.5\%$ in the skin and lungs, and $\sim 12\%$ in adipose tissues¹. Various allergens such as house dust mite, cockroach, pollen, fungi, animal hair, and food allergens directly damage epithelium which located at the interface between internal and external environment. Moreover, damage to the barrier functions of the epithelium enhances mucosal permeability of foreign substances and releasing epithelial cytokines (e.g., TSLP, IL-25, and IL-33)². Directly below the epithelium, numerous innate and acquired immune cells create a homeostasis, and ILCs act as gatekeepers to the mucosal compartment.

2. Innate Lymphoid Cells

Innate lymphoid cells (ILCs) are non-T, non-B effector cells that show high effector cell functions (mainly cytokine production) upon activation. ILCs were first described in the intestinal tract³⁻⁵, and shortly after they were also identified in the lung⁶⁻⁸. ILCs were shown to be involved with tissue homeostasis, repair and remodeling and innate immunity⁹. As non-T, non-B effector cells, ILCs lack rearranged antigen-specific receptors (TCR: T cell receptor, BCR: B cell receptor) and are therefore antigen non-specific, but they react rapidly to a wide range of innate signals. Given that ILCs produce great variety and amount of cytokines, it is surprising that ILCs had been ignored earlier. But this might be due to limited focus of immunologist on adaptive immunology, and also due to technical limits to identify several cell populations at the same time.

The classification scheme of ILCs has been developed and this scheme suggests that ILCs can be further divided into three subsets — type 1 ILCs (ILC1s and NK cells), type 2 ILCs and type 3 ILCs (ILC3s and Lti (Lymphoid Tissue inducer) cells) — based on their cytokine productions (Table 1)¹⁰. The prototypic ILC is the natural killer (NK) cell, which is called ILC1 cell or type 1 ILC, and it produces IFN- γ and expresses T-bet (T-box transcription factor)^{11,12}. The more recently discovered ILC subset is ILC2 cells or type 2 ILCs (previously called natural helper cells or nuocytes), which produce IL-5, 9 and 13 similar to the cytokines produced by Th2 cells^{3,4}. ILC2 cells require the transcription factor ROR α (retinoic acid receptor-related orphan receptor α)^{13,14} and Gata3 (GATA transcription factor)¹⁵. Type 3 ILCs or ILC3 cells have also been described with at least three different subtypes: (a) lymphoid tissue inducer (LTi) cells, which produce IL-17 and IL-22¹⁶; (b) IL-17 producing ILC3 cells, which are active in the gut of colitis patient¹⁷ and in obesity related forms of asthma⁸, and (c) IL-22 producing ILC3 cells, which are present in the skin^{18,19}, and gut²⁰.

Although ILCs have various subsets, ILC family share a common lymphoid progenitor (CLP), which is identified as Lineage-IL7R α [†]KIT⁺ ^{/low}sca-1⁺ ^{/low} and they also share common developmental pathways ^{9,14,21}. All ILCs require IL-7 signaling for survival, and Id2, a transcriptional repressor ²²⁻²⁵. These findings indicate that ILCs might be developed from a common progenitor first and then they are differentiated into different subsets based on situation. However, several studies suggested that additional pathways and plasticity might exist in the ILC lineage establishment after all ²⁵.

3. Innate lymphoid cells in allergic disease

ILCs in asthma

Asthma is one of the chronic inflammatory diseases, which affects about 300 million people worldwide²⁶. The major symptoms of asthma are shortness of breathing, coughing and wheezing caused by airwayhyperreactivity (AHR), mucus overproduction, and airway remodeling²⁷. Asthma is a heterogeneous disease which caused by many factors such as allergen exposure, virus infection, genetic background, excessive exercise and stress.

Allergic asthma was traditionally considered as T_H2-associated disease²⁸. Likewise, the discovery of ILC2 cells which produce high levels of type 2 cytokines in the lungs expands our understanding of the pathogenesis of asthma^{6,29,30}. In response to external stimuli, ILC2s accumulate in the lung and draining lymph nodes, and release large amounts of IL-5 and IL-13 which are important for the development of AHR^{31,32}. Although most studies have focused on the role of ILC2s in acute asthma, the role of ILC2 cells in chronic asthma remains to be elucidated. For example, Il1rl1^{-/-} mice had normal AHR responses after sensitization and challenge with OVA, and AHR preferentially triggered CD4⁺ Th2 cells rather than ILC2s^{33,34}. Also, ILC2s can be involved in restoration of epithelial cells by producing amphiregulin (AREG),

one of the epidermal growth factor family at the chronic phase of asthma^{35,36}. Therefore, the roles of ILC2s in the initiation of asthma versus the maintenance and resolution phases of allergic inflammation need to be investigated further. In human, ILC2s seems to contribute the development of eosinophilic asthma. The number of IL-5, IL-13 or IL-5 and IL-13 double positive ILC2s are increased in blood and sputum of patients with severe asthma than patients with mild asthma^{37,38,39}. Interestingly, submucosal expression of IL-33 was increased in children with severe therapy-resistant asthma⁴⁰ (Saglani, 2013 #296), and the numbers of ILC2s also increased in children with STRA. Therefore, these results indicated that human ILC2s also played important roles in development of asthma. However, how human ILC2s are involved in asthma pathophysiology remains to be explored.

Although most researches focus on asthma mediated by type 2 responses, non-allergic forms of asthma, triggered by environmental factors such as air pollutants (Pichavant, 2008 #295), or obesity-associated asthma appear to occur independently of T_{H2} cells⁴¹. Obesity has been shown to be a major risk factor for the development of asthma, particularly a severe, steroid-resistant form of asthma^{42,43}. Recent studies have emphasized the role type 3 cytokine in the development of asthma in both mice and humans⁴⁴⁻⁴⁶. These studies have shown that direct administration of recombinant IL-17A into the lungs can induce airway inflammation and AHR by inducing contraction of smooth muscle cells, and the source of increasing IL-17A was pulmonary Th17 cells. Moreover, the level of IL-17A increased in the sputum or peripheral blood of asthmatics and the concentration of IL-17A is correlated with the severity of asthma^{47,48}. In addition to Th17 cells, ILC3s can induce the development of AHR in obese mice⁴⁴. Obese mice developed spontaneous AHR and numbers of CCR6⁺ ILC3s producing IL-17A in the lung significantly increased. IL-1 β produced by classically activated macrophages stimulates IL-17A production from ILC3s, and blockade of IL-1R attenuate the development of obesity induced asthma⁴⁴.

All together, these studies indicate the previously unrecognized roles of ILCs in the development of various forms of asthma, and greatly expanded our knowledge of the pathogenesis of asthma.

ILCs in Atopic dermatitis

Atopic dermatitis (AD) is one of common chronic inflammatory skin diseases, which characterized by itching, dry skin, skin lesion and eczema⁴⁹. The combined factors, such as environmental factors, genetic factors, and immunological factors, can induce AD. The immune mechanisms involved in the pathology of AD has been discovered, mostly type 2 immune response is dominant in AD lesion. Th2 cells and type 2 cytokines, IL-4 and IL-13, are increased in AD skin. Moreover, IL-25, IL-33 and TSLP are also increased in AD skin and these factors promote type 2 immune responses and also modulate the skin barrier functions. Recently, it is revealed that ILC2, responding IL-25, IL-33 and TSLP, also can enhance allergic inflammation in AD⁵⁰⁻⁵². IL-33 transgenic (Tg) mouse show spontaneous phenotypes of AD at 6~8 weeks of age⁵³. In the skin lesion of Tg mouse, eosinophilic inflammation is occurred in epidermis and mast cells

are also recruited in inflamed skin. Some studies show that ILC2s in skin is more responsive to TSLP than to IL-33 and IL-25⁵⁴. Likewise IL-33 knockout and IL-25R knockout mice still develop AD. However, TSLP knockout mice, the development of AD significantly impaired. In human, there are several reports that IL-25 and IL-33 are predominant soluble factors that can enhance ILC2-mediated skin inflammation. ILC2s expresses the skin homing receptors, CCR4 and CCR10, and infiltrate into lesional skin in human ^{50,55}. Most recently, using in situ mapping of ILC in human skin, it is revealed that ILC resides epidermis near T cells, suggesting that ILC and T cells can interact and be involved in AD conditions cooperatively ⁵⁶. Moreover, AHR+ ILC3 is increased in skin biopsy of AD patients, meaning ILC3 also can be involved in AD conditions.

4. Future perspectives

Following the initial identification and characterization of ILC2s in 2010, our knowledge of this novel cell type has expanded rapidly. In mice, ILCs show both pathological and protective functions in response to various stimuli. Furthermore, ILCs network with many other immune cells, such as CD4⁺ T-cells, macrophages, eosinophils, NKT cells as well as epithelial cells, to promote the development and persistence of allergic inflammation. However, the role of ILCs in chronic and recurrent airway inflammation in asthma and other allergic diseases, such as atopic dermatitis and chronic rhinosinusitis has not been fully characterized in human. Therefore, the contributions of ILCs and molecular mechanisms underlying their activation in allergic disease should be assessed further.

References

- 1. Kim CH, Hashimoto-Hill S, Kim M. Migration and Tissue Tropism of Innate Lymphoid Cells. Trends Immunol 2016; 37:68-79.
- 2. Gon Y, Hashimoto S. Role of airway epithelial barrier dysfunction in pathogenesis of asthma. Allergol Int 2018; 67:12-7.
- 3. Moro K, Yamada T, Tanabe M, Takeuchi T, Ikawa T, Kawamoto H, et al. Innate production of T(H)2 cytokines by adipose tissue-associated c-Kit(+)Sca-1(+) lymphoid cells. Nature 2010; 463:540-4.
- 4. Neill DR, Wong SH, Bellosi A, Flynn RJ, Daly M, Langford TK, et al. Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. Nature 2010; 464:1367-70.
- 5. Buonocore S, Ahern PP, Uhlig HH, Ivanov, II, Littman DR, Maloy KJ, et al. Innate lymphoid cells drive interleukin-23-dependent innate intestinal pathology. Nature 2010; 464:1371-5.
- Chang YJ, Kim HY, Albacker LA, Baumgarth N, McKenzie AN, Smith DE, et al. Innate lymphoid cells mediate influenza-induced airway hyper-reactivity independently of adaptive immunity. Nat Immunol 2011; 12:631-8.
- 7. Monticelli L, Sonnenberg G, Abt M, Alenghat T, Ziegler C, Doering T, et al. Innate lymphoid cells promote lung-tissue homeostasis after infection with influenza virus. Nature Immunol 2011; advance online pub.
- 8. Kim H, Lee H, Chang Y-J, Pichavant M, Shore S, Fitzgerald K, et al. IL-17 producing innate lymphoid cells

- and the NLRP3 inflammasome facilitate obesity-associated airway hyperreactivity. Nature Medicine 2013:in press.
- 9. Spits H, Di Santo JP. The expanding family of innate lymphoid cells: regulators and effectors of immunity and tissue remodeling. Nat Immunol 2011; 12:21-7.
- 10. Bernink J, Mjosberg J, Spits H. Th1- and Th2-like subsets of innate lymphoid cells. Immunol Rev 2013; 252:133-8.
- 11. Strowig T, Brilot F, Munz C. Noncytotoxic functions of NK cells: direct pathogen restriction and assistance to adaptive immunity. J Immunol 2008; 180:7785-91.
- 12. Vivier E, Raulet DH, Moretta A, Caligiuri MA, Zitvogel L, Lanier LL, et al. Innate or adaptive immunity? The example of natural killer cells. Science 2011; 331:44-9.
- Halim TY, MacLaren A, Romanish MT, Gold MJ, McNagny KM, Takei F. Retinoic-acid-receptor-related orphan nuclear receptor alpha is required for natural helper cell development and allergic inflammation. Immunity 2012; 37:463-74.
- 14. Wong SH, Walker JA, Jolin HE, Drynan LF, Hams E, Camelo A, et al. Transcription factor RORalpha is critical for nuocyte development. Nat Immunol 2012; 13:229-36.
- 15. Hoyler T, Klose CS, Souabni A, Turqueti-Neves A, Pfeifer D, Rawlins EL, et al. The transcription factor GATA-3 controls cell fate and maintenance of type 2 innate lymphoid cells. Immunity 2012; 37:634-48.
- 16. Sonnenberg GF, Monticelli LA, Elloso MM, Fouser LA, Artis D. CD4(+) lymphoid tissue-inducer cells promote innate immunity in the gut. Immunity 2011; 34:122-34.
- 17. Coccia M, Harrison OJ, Schiering C, Asquith MJ, Becher B, Powrie F, et al. IL-1beta mediates chronic intestinal inflammation by promoting the accumulation of IL-17A secreting innate lymphoid cells and CD4(+) Th17 cells. J Exp Med 2012; 209:1595-609.
- 18. Villanova F, Flutter B, Tosi I, Grys K, Sreeneebus H, Perera GK, et al. Characterization of innate lymphoid cells in human skin and blood demonstrates increase of NKp44+ ILC3 in psoriasis. J Invest Dermatol 2014; 134:984-91.
- 19. Teunissen MB, Munneke JM, Bernink JH, Spuls PI, Res PC, Te Velde A, et al. Composition of Innate Lymphoid Cell Subsets in the Human Skin: Enrichment of NCR ILC3 in Lesional Skin and Blood of Psoriasis Patients. J Invest Dermatol 2014.
- 20. Tumanov AV, Koroleva EP, Guo X, Wang Y, Kruglov A, Nedospasov S, et al. Lymphotoxin controls the IL-22 protection pathway in gut innate lymphoid cells during mucosal pathogen challenge. Cell Host Microbe 2011; 10:44-53.
- 21. Zhou L. Striking similarity: GATA-3 regulates ILC2 and Th2 cells. Immunity 2012; 37:589-91.
- 22. Satoh-Takayama N, Lesjean-Pottier S, Vieira P, Sawa S, Eberl G, Vosshenrich CA, et al. IL-7 and IL-15 independently program the differentiation of intestinal CD3-NKp46+ cell subsets from Id2-dependent precursors. J Exp Med 2010; 207:273-80.
- 23. Cherrier M, Sawa S, Eberl G. Notch, Id2, and RORgammat sequentially orchestrate the fetal development of lymphoid tissue inducer cells. J Exp Med 2012; 209:729-40.
- 24. Possot C. Notch signaling is necessary for adult, but not fetal, development of ROR[gamma]t+ innate lymphoid cells. Nature Immunol. 2011; 12:949-58.
- 25. Hughes T, Briercheck EL, Freud AG, Trotta R, McClory S, Scoville SD, et al. The Transcription Factor AHR Prevents the Differentiation of a Stage 3 Innate Lymphoid Cell Subset to Natural Killer Cells. Cell Rep 2014.
- 26. Global surveillance, prevention and control of chronic respiratory diseases; A comprehensive approach. 2007.] Available from http://www.who.int/respiratory/publications/global_surveillance/en/.
- 27. Lambrecht BN, Hammad H. The immunology of asthma. Nat Immunol 2015; 16:45-56.
- 28. Robinson DS, Hamid Q, Ying S, Tsicopoulos A, Barkans J, Bentley AM, et al. Predominant Th2-like bronchoalveolar T-lymphocyte population in atopic asthma. N. Engl. J. Med. 1992; 326:298-304.
- 29. Scanlon ST, McKenzie AN. Type 2 innate lymphoid cells: new players in asthma and allergy. Curr Opin

- Immunol 2012; 24:707-12.
- 30. Kim HY, Umetsu DT, Dekruyff RH. Innate lymphoid cells in asthma: Will they take your breath away? Eur J Immunol 2016; 46:795-806.
- 31. Wolterink RG, KleinJan A, van Nimwegen M, Bergen I, de Bruijn M, Levani Y, et al. Pulmonary innate lymphoid cells are major producers of IL 5 and IL 13 in murine models of allergic asthma. European journal of immunology 2012; 42:1106-16.
- 32. Halim TY, Krauß RH, Sun AC, Takei F. Lung natural helper cells are a critical source of Th2 cell-type cytokines in protease allergen-induced airway inflammation. Immunity 2012; 36:451-63.
- 33. Guo L, Huang Y, Chen X, Hu-Li J, Urban JF, Jr., Paul WE. Innate immunological function of TH2 cells in vivo. Nat Immunol 2015; 16:1051-9.
- 34. Endo Y, Hirahara K, Iinuma T, Shinoda K, Tumes DJ, Asou HK, et al. The interleukin-33-p38 kinase axis confers memory T helper 2 cell pathogenicity in the airway. Immunity 2015; 42:294-308.
- 35. Monticelli LA, Sonnenberg GF, Abt MC, Alenghat T, Ziegler CG, Doering TA, et al. Innate lymphoid cells promote lung-tissue homeostasis after infection with influenza virus. Nat Immunol 2011; 12:1045-54.
- 36. Wills-Karp M, Finkelman FD. Innate lymphoid cells wield a double-edged sword: type 2 cytokine-producing innate lymphoid cells are present in human and mouse lungs, where they contribute to both type 2 immune responses and tissue repair. Nature immunology 2011; 12:1025-8.
- 37. Liu T, Wu J, Zhao J, Wang J, Zhang Y, Liu L, et al. Type 2 innate lymphoid cells: A novel biomarker of eosinophilic airway inflammation in patients with mild to moderate asthma. Respir Med 2015; 109:1391-6.
- 38. Smith SG, Chen R, Kjarsgaard M, Huang C, Oliveria JP, O'Byrne PM, et al. Increased numbers of activated group 2 innate lymphoid cells in the airways of patients with severe asthma and persistent airway eosinophilia. J Allergy Clin Immunol 2016; 137:75-86 e8.
- 39. Nagakumar P, Denney L, Fleming L, Bush A, Lloyd CM, Saglani S. Type 2 innate lymphoid cells in induced sputum from children with severe asthma. J Allergy Clin Immunol 2016; 137:624-6 e6.
- 40. Muller U, Stenzel W, Kohler G, Werner C, Polte T, Hansen G, et al. IL-13 induces disease-promoting type 2 cytokines, alternatively activated macrophages and allergic inflammation during pulmonary infection of mice with Cryptococcus neoformans. J Immunol 2007; 179:5367-77.
- 41. Kim HY, DeKruyff RH, Umetsu DT. The many paths to asthma: phenotype shaped by innate and adaptive immunity. Nat Immunol 2010; 11:577-84.
- 42. Holguin F, Bleecker ER, Busse WW, Calhoun WJ, Castro M, Erzurum SC, et al. Obesity and asthma: an association modified by age of asthma onset. J Allergy Clin Immunol 2011; 127:1486-93 e2.
- 43. Camargo CA, Jr., Weiss ST, Zhang S, Willett WC, Speizer FE. Prospective study of body mass index, weight change, and risk of adult-onset asthma in women. Arch Intern Med 1999; 159:2582-8.
- 44. Kim HY, Lee HJ, Chang YJ, Pichavant M, Shore SA, Fitzgerald KA, et al. Interleukin-17-producing innate lymphoid cells and the NLRP3 inflammasome facilitate obesity-associated airway hyperreactivity. Nat Med 2014; 20:54-61.
- 45. McKinley L, Alcorn JF, Peterson A, Dupont RB, Kapadia S, Logar A, et al. TH17 cells mediate steroid-resistant airway inflammation and airway hyperresponsiveness in mice. J Immunol 2008; 181:4089-97.
- 46. Kudo M, Melton AC, Chen C, Engler MB, Huang KE, Ren X, et al. IL-17A produced by alphabeta T cells drives airway hyper-responsiveness in mice and enhances mouse and human airway smooth muscle contraction. Nat Med 2012; 18:547-54.
- 47. Sun YC, Zhou QT, Yao WZ. Sputum interleukin-17 is increased and associated with airway neutrophilia in patients with severe asthma. Chin Med J (Engl) 2005; 118:953-6.
- 48. Agache I, Ciobanu C, Agache C, Anghel M. Increased serum IL-17 is an independent risk factor for severe asthma. Respir Med 2010; 104:1131-7.
- 49. Peng W, Novak N. Pathogenesis of atopic dermatitis. Clin Exp Allergy 2015; 45:566-74.
- 50. Salimi M, Barlow JL, Saunders SP, Xue L, Gutowska-Owsiak D, Wang X, et al. A role for IL-25 and

- IL-33-driven type-2 innate lymphoid cells in atopic dermatitis. J Exp Med 2013; 210:2939-50.
- 51. Deleuran M, Hvid M, Kemp K, Christensen GB, Deleuran B, Vestergaard C. IL-25 induces both inflammation and skin barrier dysfunction in atopic dermatitis. Chem Immunol Allergy 2012; 96:45-9.
- 52. Kim BS. Innate lymphoid cells in the skin. J Invest Dermatol 2015; 135:673-8.
- 53. Imai Y, Yasuda K, Sakaguchi Y, Haneda T, Mizutani H, Yoshimoto T, et al. Skin-specific expression of IL-33 activates group 2 innate lymphoid cells and elicits atopic dermatitis-like inflammation in mice. Proc Natl Acad Sci U S A 2013; 110:13921-6.
- 54. Kim BS, Siracusa MC, Saenz SA, Noti M, Monticelli LA, Sonnenberg GF, et al. TSLP elicits IL-33-independent innate lymphoid cell responses to promote skin inflammation. Sci Transl Med 2013; 5:170ra16.
- 55. Yang J, Hu S, Zhao L, Kaplan DH, Perdew GH, Xiong N. Selective programming of CCR10(+) innate lymphoid cells in skin-draining lymph nodes for cutaneous homeostatic regulation. Nat Immunol 2016; 17:48-56.
- 56. Bruggen MC, Bauer WM, Reininger B, Clim E, Captarencu C, Steiner GE, et al. In Situ Mapping of Innate Lymphoid Cells in Human Skin: Evidence for Remarkable Differences between Normal and Inflamed Skin. J Invest Dermatol 2016; 136:2396-405.